Development of Modified Instant Starch from Taro (Colocasia esculenta) by Gelatinization

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Abstract: In present study an attempt to isolate the starch from Taro (Colocasia esculenta) by adopting standard wet milling extraction process. Isolated starch was then pregelatinized to convert into seven samples of instant starch by autoclaving for different duration ranging from 1hr. to 4hr. with 30 minutes increment span. The seven samples of instant starch so prepared were designated as IS-1, IS-1.5, IS-2, IS-2.5, IS-3, IS-3.5 and IS-4 and isolated taro starch was used as control sample (IS 0). Starch after gelatinization was dried and packed in polyethylene bag. Isolated starch and instant starch samples were analyzed for; resistant starch content (RS %), amylose content (%), water absorption capacity (g/g), swelling power (g/g), solubility (g/g), enzyme digestibility (ED %) and dispersibility (%). The functional parameters of Instant starch were compared with control starch (IS-0). Result showed that the RS content of IS-3.0 was reported to be increased with the span of autoclaving and 3hr was found to be optimum time for making Instant starch with %RS(30.14). Other parameters of IS3, were % amylose content (17.21), swelling power (18.28g/g), solubility (0.099g/g), water absorption capacity (1.72g/g), % enzyme digestibility (59.32) and % dispersibility (68), while for isolated untreated starch (IS-0) it were amylose content % (17.83), swelling power (16.02), solubility (0.098), water absorption capacity (1.64), % enzyme digestibility (56.54) and % dispersibility (83). Data reveal that functional properties including (water absorption capacity, swelling power, solubility, and enzyme digestibility) were improved in IS3.0. It was reported that amylose content and dispersibility was slightly reduced on autoclaving owing to degradation and retrogradation of starch. Study concludes that instant taro starch can be prepared by pregelatinizing the starch for 3hr at 15 psi with improved water absorption capacity, swelling power, solubility, enzyme digestibility compared to control starch. Cake prepared from optimum 20% level of IS-3 starch resulted in cake with better volume and texture.

Keywords: Amylose content, instant starch, pregelatinization, resistant starch, retrogradation.

I. Introduction

Starch is a carbohydrate consisting of a large number of glucose units joined together by glycosidic bonds. This polysaccharide is produced by all green plants as an energy store. It is the most common carbohydrate in the human diet and is contained in large amounts in such staple foods as potatoes, wheat, maize (corn), rice, and cassava. Depending on the plant, starch contains 20 to 25% amylose and 75 to 80% amylopectin [1].

In the early days, starch was considered to be completely digested and absorbed in the small intestine. Later, it was found that a fraction of starch (resistant starch or RS) survived the enzyme digestion in the digestive track [2]. RS is not absorbed in the small intestine, and is passed onto the colon, where it is fermented by the gut microflora. The fermentation of RS in the colon produces short-chain fatty acids and other organic acids [3, 4, 5], and releases hydrogen through exhalation [6].

RS has demonstrated similar physiological benefits as dietary fibers [7]. and it has been proposed that RS should be included in the definition of dietary fibers [8, 9]. Consumption of RS has shown to reduce glucose absorption and insulin secretion, which prevents insulin resistance and metabolic syndrome, including obesity, diabetes, and heart disease [10, 11]. Butyrate, a product of RS fermentation in the colon can prevent the development of cancerous cell in the colon [12, 13]. Furthermore, the amount of RS that reaches the colon is larger than that of other dietary fibers, thus it is more important as a substrate for colonic fermentation [14]. Examples of RS include raw potato starch, raw high-amylose maize starch, raw green banana starch, and retrograded amylose in bread [15].

RS has been classified into four types according to the nature of the enzyme resistance and the structure of the starch [16, 17]. RS type 1 (RS1) is physically inaccessible starch, which is protected by a protein matrix or cell wall material, such as that in whole grains, legumes, and pasta. RS type 2 (RS2) is native, uncooked semi-crystalline granular starch that displays the B- and some C-type polymorphs, such as uncooked potato starch, green banana starch, and high-amylose maize starch. RS type 3 (RS3) is retrograded amylose formed in cooked starchy food. RS type 4 (RS4) is chemically modified or cross-linked starch, which is less accessible for

enzyme hydrolysis. In addition to the four types of RS, resistant dextrins [18, 19] and amylose-lipid complexes [20, 21] are also included in RS because they are resistant to enzyme hydrolysis.

Study conducted to determine the nutritional fact of Resistant Starch (RS). RS in food products resists the in vitro enzyme hydrolysis as well as remains undigested in human small intestine. However RS reaches to colon it's being fermented with production of volatile short chain fatty acid. According to literature and present study it may be summarized the development of RS is due to retrogradation of starch and cooling of gelatinized starch. Starch gelatinization is quick process so that developed gelatinized starch can be termed as Instant Starch (IS). Motive of the study is to develop the Instant starch with high RS and improved functional properties Viz. amylose content, water absorption capacity, swelling power, solubility, enzyme digestibility (ED) and dispersibility.

II. Materials and Methods

Fresh Taro tubers were purchased from local market of Jalgaon of Varity C. esculenta var. globulifera Engl **2.1 Extraction of Starch from Tubers**

Taro tubers was collected and cleaned by proper washing. After washing of tubers outer skin was removed by peeling. The tubers ware then sliced and chopped and kept for drying in hot air oven at 55 ^oC. The dried tuber pieces were then crushed in mixer grinder to form the powder. This powder is further used for isolation of starch. Starch was isolated by standard wet milling process

Taro starch was isolated according to the procedures used by Azhar Ahmed, Farukh Khan (2013) [22]. Taro powder (50 gm) was dispersed in a sodium metabisulphite solution (150 ml, 0.45% W/V) for 12 hours at refrigeration temperature. Now the slurry is was milled using a laboratory food blender for 5 min. The blended slurry was mixed with 450 ml NaCl solution (0.1 M) and 50 ml Toluene. The mixture was agitated for 1hr and then allowed to stand until the starch granules precipitate at the bottom. The protein in toluene and NaCl solution layers was siphoned off and discarded, and this was repeated until all of the proteins were removed as indicted by a clear toluene layer after the starch granules settled at the bottom. The starch layer was then washed with water several times and subsequently with absolute ethanol. The wet milled starch was then recovered by filtration through Whatman filter paper, rinsed with ethanol and air dried.

2.2 Instant starch Preparation (Gelatinization of starch):

Autoclaving of Starch:

Starch gelatinization was performed using the method of Shamai et al. (2003) and Huth et al. (2000) [23, 24], with a slight modification. For each treatment, taro starch (180 g on a dry basis) was weighed in a 1,000mL flask before 720 ml distilled water was added (starch: water = 1:4, w/v). The flask was then kept on magnetic stirrer for stirring at ambient temperature for 30 min, then covered with aluminum foil and heated by laboratory autoclave for different duration ranging from 1hour to 4hour with 30 minutes increment span at 121°C. After each treatment, the starch gel was cooled at ambient temperature for 2 hours and stored in polythene bags for analysis.

Physicochemical analysis Method and instrument.			
Moisture (%)	S. Ranganna1995, Hot air oven		
Ash (%)	S. Ranganna1995, Muffle furnace		
Fat (%)	S. Ranganna1995, solvent extraction method		
Protein (%)	S. Ranganna1995, Protein analyzer (kjaldhal)		
Crude fiber (%)	S. Ranganna1995, Crude fiber analyzer		
Carbohydrates (%)	S. Ranganna1995[25]., UV-Double Beam Spectrophotometer 2205		
RS content (%)	Onyango et al. (2006) [26], laboratory centrifuge REMI-R4C		
Amylose content (%)	Morrison and Laignelet (1983) [27], UV-VIS Spectrophotometer		
Swelling power (g/g)	Jirarat Tattiyakul et al (2006) [28], laboratory centrifuge REMI-R4C		
Solubility (g/g)			
WAC (g/g)	Medcalf and Gilles (1965) [29], laboratory centrifuge REMI-R4C		
ED (%)	S. Ranganna1995, Incubator.		
Dispersibility (%)	AACC 56-61A, 2000b [30].measuring cylinder		

Table 2 1. Methods for the Physicochemical Analysis and Instruments

2.3 Characterization of Isolated starch and instant starch.

2.4 Fourier transforms infrared spectroscopy (FTIR)

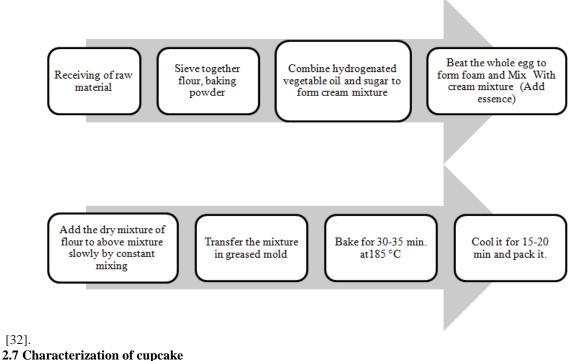
Fourier transform infrared spectra were recorded using a Jasco FTIR spectrometer. Starch Particles were collected using the KBr pellet method. A total of 32 scans were Obtained and the resolution was 4 cm-1. The wavelength region was between 4000 and 400 cm-1. All spectra were baseline corrected and normalized through setting the maximum transmittance to 100%. [31].

2.5 Granule microscopy:

Starch granule shapes were observed and photographed using a BX 50 microscope. Granule shape was measured using a microscope fitted with a calibrated eyepiece.

2.6 Cup cake preparation

[32].



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Physicochemical analysis	Method and instrument.					
Cake Volume	AACC 10-10.03, (2000).					
Density	AACC 10-10.03, (2000). [33].					

Result And Discussion III.

Table 3.1: Results of the Compositions of Taro tubers	
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Tuble 5.1. Results of the compositions of Tub tubers				
Parameter	Gram %			
Moisture	72.5±0.73			
Ash	0.94±0.01			
Fat	0.26±0.01			
Protein	2.45±0.08			
Crude fiber	1.21±0.02			
Carbohydrates	22.64±0.73			

Table3.1 showed chemical composition of Taro Tubers of verity C. esculenta var. globulifera Engl Proximate compositions of physical parameters was in between the standard literature values results showed that the parameters were moisture 72.5 %, Ash 0.94 %, fat 0.26 % And protein 2.45 % but the crude fiber content of taro tuber w as comparatively higher than that of the literature value (1.21%).

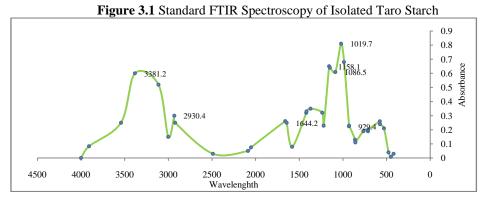


Figure 3.1 showed the fingerprint region of the spectrum, three characteristic peaks appear between 929 and 1158/cm, which are attributed to C–O bond stretching. The peaks at 1086 and 1019/cm are characteristic of O–C stretching associated with the anhydroglucose ring. Peak near 850 cm–1 corresponded to the C–H deformations another characteristic peak occurs at 1644/cm, which may be related to the presence of tightly bound water in the starch. The peaks appearing at 3381–3800 /cm in the spectrum were due to hydrogen-bonded hydroxyl groups (O-H), which is attributable to the complex vibrational stretches associated with free, inter-, and intra-molecular bound hydroxyl groups, which make up the gross structure of starch. The sharp band at 2930/cm is characteristic of C–H stretching associated with the ring methane hydrogen atoms.

Figure 3.2: light Microscopic micrograph of taro starch and potato starch granules (isolated) in normal and zoom

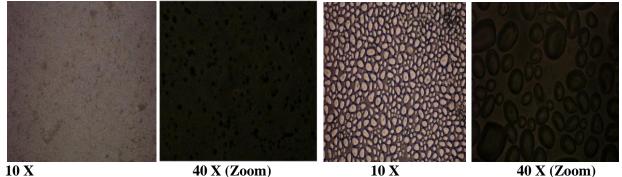
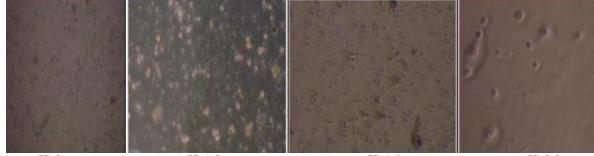


Figure 3.2 show that the starch granule shapes of potato and taro starches. They were present in different shapes. Light microscopy of starch granules from taro starches as compared with potato starches. The mean dimension of starch granule size measured by microscopy of potato starch was higher than that of taro starch. The particle size distribution of Taro starch was the most homogeneous, while that of potato starch was the most free and separated than Taro Starch. Granule size and particle size distribution are characteristics that markedly influence the functional properties of starch granules [34]. The size range of the taro starch was clearly different from that of potato starch. The granule of taro starch is very small.

3.1 RS content

Figure 3.3: light microscopic micrograph of Instant Taro starch 121°C.

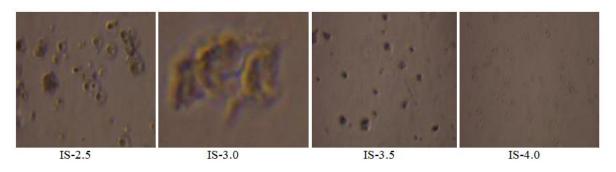


IS-0

IS-1.0

IS-1.5

IS-2.0



Autoclaving	Resistant starch (%)
00 hr (IS-00)	09.82±0.18
1.0hr (IS-01)	23.22±1.02
1.5hr (IS-1.5)	26.35±1.37
2.0hr (IS-2.0)	28.72±1.16
2.5hr (IS-2.5)	29.31±1.05
3.0hr (IS-3.0)	30.14±1.06
3.5hr (IS-3.5)	29.13±0.95
4.0hr (IS-4.0)	28.36±0.81

Table3.2: Resistant starch content at differen	t autoclaving time
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Figure 3.3 shows that the starch granule shapes of native taro starch and after modification 121°C 1hr, 1.5hr, 2hr, 2.5hr, 3hr, 3.5hr and 4hr. Isolated starch granules are oval shape without any hollow area inside and very fine distribution. The hollow area inside the starch granules in IS-1 (1hr) starch was observed but the shapes of the IS-1 (1hr) starch granules are contorted to a folded structure.

Microscopic images showed that gelatinization of starch changed the shape of the starch granules to a folded structure. Folding was highly observed in 2.5 hr and 3 hrs autoclaving. In both 2.5 hr and 3 hr autoclaving the resistant starch content is also comparatively high. In 3.5hr and 4hr autoclaving resistant starch content is lesser than 2.5 hr and 3 hr autoclaving.

From this microscopic picture it was well established that starches were modified

Parameters	Isolated Starch	Instant starch (IS 3.0)
Amylose content (%)	17.83±0.01	17.21±0.00
Swelling power (g/g)	16.02±0.01	18.28±0.01
Solubility (g/g)	0.098 ± 0.00	0.099±0.00
WAC (g/g)	1.64±0.01	1.72±0.01
ED (%)	59.32±0.28	56.54±0.23
Dispersibility (%)	83±1.22	68±0.82

Table 3.3: Results of the Physicochemical Analysis of Isolated starch and Instant starch (IS-3)

Results of the physicochemical analysis of isolated and IS 3.0 are shown in Table 3.3 as per the result obtained from the analysis. There was 3hr autoclaving resulted in improvement of resistant starch content (30.14%) & functional properties Viz. Swelling power (18.28 g/g) and solubility (0.099g/g) which was higher than that of isolated starch Swelling power (16.02 g/g) and solubility (0.098g/g) While dispersibility of IS was quietly reduced it may be due to retrogradation on storage. There was no significant change in the water absorption capacity, enzyme digestibility and amylose content.

Ingredients	CAKE0	CIS-05	CIS-10	CIS-15	CIS-20	CIS -25	CIS -30	CIS -45
Sugar	100	100	100	100	100	100	100	100
Flour	100	95	90	85	80	75	70	55
HMTS (RS)	00	05	10	15	20	25	30	45
Eggs	2	2	2	2	2	2	2	2
Baking powder	6.3	6.3	6.3	6.3	6.3	6.3	6.3	6.3
Fat (Butter)	40	40	40	40	40	40	40	40
Milk	80	80	80	80	80	80	80	80
Condensed Milk	15	15	15	15	15	15	15	15
Salt	3.0	3.0	3.0	3.0	3.0	3.0	3.0	3.0
Vanilla	2.5	2.5	2.5	2.5	2.5	2.5	2.5	2.5

Table3.4: Instant starch incorporated cup cake formulations

Cake0 (Control): cup cake prepared by wheat flour only.

CIS -5: cup cake prepared by replacing the flour with 5% Instant starch.

CIS -10: cup cake prepared by replacing the flour with 10% Instant starch.

CIS -15: cup cake prepared by replacing the flour with 15% Instant starch.

CIS -20: cup cake prepared by replacing the flour with 20% Instant starch.

CIS -25: cup cake prepared by replacing the flour with 25% Instant starch.

CIS -30: cup cake prepared by replacing the flour with 30% Instant starch.

CIS -45: cup cake prepared by replacing the flour with 45% Instant starch

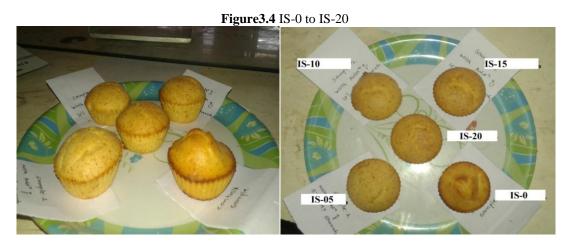
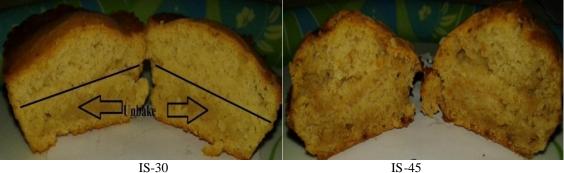
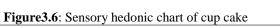


Figure3.5: Cup cakes with IS-0 IS-5, IS-10, IS-15, and IS-20







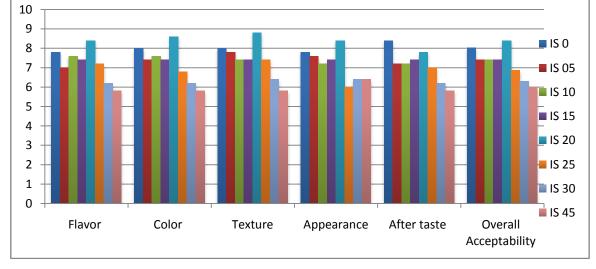


Figure 3.6 shows in sample HMTS-5, HMTS-10, HMTS-15 and HMTS-20 successful replacement of HMTS with flour has been done. Properties like texture, sponginess and mouth feel were improved. In sample HMTS-45 there were all the quality parameters are collapse down and such product is unfit for consumption.

In sample HMTS-30 the product is not properly baked. Smoothness and sponginess are not palatable. In HMTS-25 sponginess, volume of cake, texture and other quality parameter are good but slightly product remains unbaked. So it is conceders as unacceptable.

So all the above results shows the best formulation is in CIS-20. It gives grater loaf volume, sponginess, texture, color and other sensory qualities than control sample.

Tuble 5.6. Thysicoencinical parameters of Cib 20					
Parameter	CAKE0	CIS-20			
Weight (g)	37.38±0.73	36.71±0.56			
Volume (cm ³)	111.45±1.40	116.16±1.88			
Moisture (%)	15.63±1.31	14.21±1.06			
Ash (%)	1.46±0.18	1.21±0.24			
Fat (%)	17.86±0.98	18.23±1.27			
Protein (%)	9.84±0.75	7.78±0.98			

 Table 3.5: Physicochemical parameters of CIS-20

Table3.5 showed chemical composition of the produced cake substitutes CIS-20. Moisture content cake of CIS-20 was comparatively lesser than control sample. CIS-20 Cake showed significant differences for total fat and ash content relative to control cake and this may be due to the higher content of total fats and ash in CIS. Protein of CISS20 substituted cake decreased significantly compared with control cake. This may be due to replacement of wheat flour with starch substitution.

Physical attributes (weight, volume, specific volume and density) of wheat flour cake and CIS-20 cake are given in Table 3.5 According to weight data showed no much difference between Cake-0 (37.38) and CIS-20 (36.71). CIS-20 had comparatively high volume (116.16 cm³), than control cake (111.45cm³). Results had been showed that Instant starch in cake improves the loaf volume, Texture and sponginess of cake rather increases in weight.

IV. Conclusion

Study concludes that instant starch prepared by 3hr autoclaving resulted in improvement of resistant starch content (30.14%) & functional properties Viz. Swelling power (18.28 g/g) and solubility (0.099g/g) which was higher than that of isolated starch Swelling power (16.02 g/g) and solubility (0.098g/g). It was also noticed that the 20% replacement of Instant starch resulted in improvement in the cup cake quality which might be due to high swelling or gas retention capacity of instant starch.

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